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Expression analyses of insulin-like peptide 3, RXFP2, LH receptor and
3β-HSD in testes of normal and cryptorchid dogs

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Abstract

Insulin-like peptide 3 (INSL3) plays a key role in testicular descent in rodents, whereas in domestic animals, many aspects of the roles of INSL3 in reproductive organs after puberty are still unknown. This study was undertaken to: (1) determine the quantitative changes of gene expression of testicular INSL3, its receptor (RXFP2), LH receptor and 3β-HSD during and after puberty in normal male dogs; (2) compare the expressions of these substances in normal and cryptorchid dogs; and (3) localize the cells expressing INSL3 in normal and retained canine testes. Testes were obtained from small-breed normal male dogs (n=56) and cryptorchid dogs (n=22). Normal scrotal testes from the normal dogs (normal testes) and retained testes from both the unilateral and bilateral cryptorchid dogs (retained testes) and scrotal testes of the unilateral cryptorchid dogs (cryptorchid scrotal testes) were used. We measured the concentrations of these testicular mRNAs by quantitative real-time RT-PCR, and an enzyme immunoassay was used for measuring INSL3 peptide. Immunohistochemistry for INSL3 peptide was done in paraformaldehyde-fixed frozen testicular tissue. In the normal dogs, total amount of INSL3 mRNA per testis tended to decrease (P=0.05) from pubertal (6–12 mo) to post-pubertal (1–5 y) and decreased (P<0.01) to middle age (5–10 y), but total amount of INSL3 peptide per testis did not change among age groups. Concentrations of INSL3
mRNA were higher (P<0.01) in retained testes than those in the normal testes and cryptorchid scrotal testes, and similar differences were observed for INSL3 peptide.

Reversely, total amounts of INSL3 mRNA and peptide per retained testis were lower (P<0.01) than those per normal testis, due to smaller weight of retained testes.

Concentrations and total amount of RXFP2 mRNA in the retained testes were almost nil, and lower (P<0.01) than those in the normal testes and in the cryptorchid scrotal testes. Total amount of LH receptor mRNA per retained testis was lower (P<0.01) than that per normal testis. The immunohistochemical analysis revealed that INSL3 was expressed only in Leydig cells of both the normal and retained canine testes. These results suggest that INSL3 in retained testes of cryptorchid dogs is substantially expressed per unit-weight basis, but may be produced with lower amount as a whole testis. Also this study provides findings that RXFP2 gene is expressed scarcely in the retained testes, but normally in cryptorchid scrotal testes.

**Keywords:** INSL3; RXFP2; Leydig cell; Cryptorchid; Testis; Dog
1. Introduction

Insulin-like peptide 3 (INSL3), also known as relaxin-like factor, is a relatively newly identified peptide hormone produced by testicular Leydig cells [1–3]. Its mRNA is constitutively expressed in a differentiation-dependent manner related to the postnatal development of Leydig cell function [1, 4]. During the fetal period, INSL3 plays an important role in the trans-abdominal phase of testicular descent in mice [5, 6] and the survival of germ cells as an anti-apoptotic factor in adult humans [7] and rats [8]. INSL3 has also been suggested to have an important endocrine role in the males of many mammalian species and can readily be measured in the peripheral plasma of humans [9–11], rodents [12, 13], cattle [14] and dogs [15].

In male dogs, plasma INSL3 concentrations increased significantly from pre-pubertal to pubertal age and then declined from pubertal to post-pubertal age [15]. Lower INSL3 concentrations have been detected in bilateral cryptorchid dogs compared to normal and unilateral cryptorchid dogs, suggesting the diagnostic value of this hormone in anticipating bilaterally retained testes [15]. However, the dynamics of the expression of INSL3 at the mRNA and peptide levels associated with the development of reproductive stages remain to be elucidated.

Relaxin family peptide receptor 2 (RXFP2; formerly known as LGR8) is the
specific receptor of INSL3 [16]. RXFP2 knockout mice showed intra-abdominal cryptorchidism and male infertility due to the arrest of spermatogenesis [17, 18]. The expression of RXFP2 in adult testes was demonstrated to be localized in germ cells in seminiferous tubules and interstitial Leydig cells in humans [7], rats [7, 19] and mice [7, 20] by reverse transcription-polymerase chain reaction (RT-PCR) and immunohistochemistry. In dogs, INSL3 and RXFP2 expression were revealed by immunohistochemistry in testicular Leydig cells of both normal and cryptorchid testes, with a lack of RXFP2 expression in the genital tracts of cryptorchid testes [21]. The quantitative changes of this receptor during sexual development in canine testes have not yet been determined.

Cryptorchidism, a failure of one or both testes to descend normally into the scrotum, affects 2%–9% of newborn boys [22], 2%–8% of male horses [23] and 1.2%–10.7% of male dogs [24, 25], with a higher risk in small breeds than in larger breeds dogs [26]. INSL3/RXFP2 signaling plays a crucial role in the process of testicular descent in mice, but differences in the testicular expressions of INSL3 and RXFP2 between normal and cryptorchid animals have not been analyzed quantitatively in any species including dog, to the best of our knowledge.

LH receptor and 3β-hydroxysteroid dehydrogenase (3β-HSD; a steroidogenic
enzyme) have also been used as a marker of testicular Leydig cells and have been identified in horses [27], rats [28] and dogs [29]. These markers were used for identifying normal and tumorous Leydig cells in dogs [29]. In equine testes, the immuno-labeling of 3β-HSD was very weak or absent in immature Leydig cells of pre-pubertal testes and increased in post-pubertal and adult testes [30]. Steroidogenesis occurs primarily in Leydig cells [31], and reduced testosterone production has been observed in cryptorchid mice, stallions and dogs [32–34]. To the best of our knowledge, there have been no studies comparing the LH receptor and 3β-HSD gene expressions among normal, scrotal and retained testes in dogs.

The objectives of the present study were to: (1) determine the quantitative changes of the gene expressions of testicular INSL3, RXFP2, LH receptor and 3β-HSD during and after puberty in normal male dogs; (2) compare the expressions of these substances in retained and scrotal testes of cryptorchid dogs with those of normal testes of normal dogs; and (3) localize cells expressing INSL3 in normal and cryptorchid canine testes.

2. Materials and methods

2.1. Animals and sampling
A total of 78 male dogs were used in the present study. The dogs were presented to a private animal clinic close to our university for ordinary contraception or treatment of cryptorchidism. All of the dogs were privately owned, and the owners’ consent was obtained before the collection of samples. The study was conducted according to the regulations of the local Institutional Animal Care and Use Committee. Before surgery, testicular presence was checked manually and diagnosed as normal (n=56) if both testes were palpable inside the scrotum. Cryptorchidism was diagnosed (n=22) when one (unilateral, n=16) or both (bilateral, n=6) testes were missing in the scrotum after 6 months of age [35, 36]. All dogs belonged to small breeds, and nearly 80% were Toy Poodles, Chihuahuas, Miniature Dachshunds, Pomeranian and Shih Tzus. The ages of the dogs ranged from 6 mo to 10 y. The range of body weights was 1.4 to 8.6 kg (4.4 ± 0.2 kg; mean ± SEM). Testes samples were collected after castration or cryptorchidectomy and then immediately dispatched to the laboratory on ice. The testes were separated from the epididymides. The weight of both testes was recorded from all normal and cryptorchid dogs.

2.2. Tissue processing

Different testes samples were used for (1) a quantitative RT-PCR and enzyme
immunoassay (EIA) and (2) immunohistochemistry. Normal testes (either the right or left testis) of the normal dogs were used for the RT-PCR and EIA (n=46) and for immunohistochemistry (n=10). Retained testes of the unilateral and bilateral cryptorchid dogs were used for the RT-PCR and EIA (n=19) and for immunohistochemistry (n=5). Scrotal testes of the unilateral cryptorchid dogs (cryptorchid scrotal testes) were used only for the RT-PCR and EIA (n=11). For the quantitative RT-PCR and EIA, testicular tissue was cut into small pieces (approx. 1 cm³) and saved at −80°C until RNA and peptide extractions.

For the immunohistochemistry, testicular tissues were fixed overnight in 4% paraformaldehyde, followed by incubating in sucrose solutions (10%, 20% and 30%) for an additional 24 h at 4°C. The tissue pieces were then embedded in OCT compound (Tissue-Tek, Sakura Finetek Japan, Tokyo) and maintained at −80°C until sectioning.

2.3. RNA extraction, cDNA synthesis and real-time PCR

Total RNA was isolated from a small amount of frozen testicular tissue (approx. 20 mg) using the RNeasy Mini Kit (QIAGEN, Hilden, Germany), according to the manufacturer’s instructions. RNA quantity and quality were evaluated using a spectrophotometer (U-2000, Hitachi, Tokyo) at 260 nm. The isolated total RNA was
stored at $-80^\circ$C until RT-PCR.

Table 1 lists the pairs of primers used to quantitate mRNAs for canine INSL3, RXFP2, LH receptor, 3β-HSD and 18S rRNA in testicular tissue and the expected sizes of their base pairs. The primers other than 18S rRNA were designed based on the canine nucleotide sequence registered in GenBank. The 18S rRNA primers were used as an internal standard as reported [37].

The mRNAs were measured by reverse transcription and quantitative real-time PCR with calibration curves. For the calibration of cDNA for each targeted mRNA from the total RNA, an ordinary RT-PCR was performed with a Takara RNA PCR Kit (AMV) Ver. 2 (Takara, Ohtsu, Japan) according to the manufacturer’s instructions. The PCR products were stored at $-20^\circ$C until these analyses. A portion of the PCR products was electrophoresed through a 2.0% agarose gel containing 0.5 mg/mL ethidium bromide. The band was dissected out on an UV transilluminator, and DNA was extracted from the agarose gel, using a QIAEX II Extraction Kit (QIAGEN, Hilden, Germany). Purified PCR products were sequenced directly using a sequencer (3730xl DNA Analyzer, Applied Biosystems, Carlsbad, CA) by outsourcing (Bio Matrix Research, Chiba, Japan).

The cDNA sequence data were compared with the registered sequences in
GenBank using sequence analysis software (Sequence Scanner, Applied Biosystems).

All five targeted cDNA sequences were identical at 100% with the registered sequences in GenBank. The registered cDNA sequences in GenBank for INSL3, RXFP2, LH receptor, 3β-HSD and 18S rRNA are NM_001002962, NM_001005870, XM_538486, NM_001010954 and NR_046237, respectively.

The total RNA (0.5 μg) from canine testes was reverse-transcribed into cDNA using the iScript™ Select cDNA Synthesis Kit (Bio-Rad Laboratories, Hercules, CA) according to the manufacturer’s instructions. The reverse transcription was reacted in a Real-Time PCR System (Bio-Rad Laboratories). The subsequent real-time PCR reaction was performed using Ssofast™ EvaGreen® Supermix (Bio-Rad Laboratories) per the manufacturer’s instructions. The numbers of cycle for the PCR reactions of INSL3, RXFP2, LH receptor, 3β-HSD and 18S rRNA were 40, 40, 34, 34 and 20, respectively. The concentrations of mRNA were calculated as the threshold cycle numbers of targeted mRNA for each sample divided by those of 18S rRNA. Total amount of targeted mRNA per testis was calculated from the data of mRNA concentration and testicular weight. The standards were checked for linearity in every assay with serial 10-fold diluted calibration cDNA for each targeted mRNA. The regression coefficient (R²) value was more than 0.996 in all assays.
2.4. Extraction of INSL3 from testicular tissue

The extraction of INSL3 from testicular tissue was carried out according to the procedure described earlier for bovine plasma in our laboratory [14]. First, approx. 100 mg of frozen testicular tissue was placed into a tube containing 500 μL of 0.1% trifluoroacetic acid (TFA). Homogenization was then performed for 1 min (20 s × 3) on ice using a Polytron homogenizer (Kinematica, Littau, Switzerland). Another tube with 300 μL of acetonitrile was kept ready in advance, into which 500 μL of the homogenized mixture was transferred immediately after homogenization. This was then kept at 4°C for overnight after mixing by vortexing. Next, the mixture was centrifuged at 15,000 × g for 10 min at 4°C. The resulting supernatant was then transferred into another tube and concentrated by a vacuum centrifugation (Centrifugal Concentrator CC-105; Tomy Seiko, Tokyo) for approx. 3 h (final volume, approx. 60 μL). Finally, 450 μL of 0.05 M phosphate buffer (pH 7.5) was added to the concentrated supernatant, which was stored at −30°C until the assay.

2.6. INSL3 assay

The concentrations of INSL3 peptide were determined using an EIA. The
immunoassay procedure was basically similar to the previously described time-resolved fluorescence immunoassay (TRFIA) [15], except that biotinylated canine INSL3 was used for the EIA instead of europium-labeled human INSL3. Briefly, eight-well strips were coated with 100 µL of anti-mouse IgG antibody (MP Biochemicals, Solon, OH; 5 µg/mL in 0.05M sodium bicarbonate; pH 9.7), and nonspecific binding sites were blocked overnight with assay buffer containing 2% bovine serum albumin (BSA; Cohn Fraction V, Sigma-Aldrich, St. Louis, MO), and 0.02% ProClin 950 (Sigma-Aldrich) in 0.01M PBS, pH 7.4.

Next, 50 µL of canine INSL3 standard [15] or sample medium and 50 µL of anti-bovine INSL3 mouse monoclonal antibody (2-8F [14, 15]; 1:1,000,000 dilution in assay buffer) were dispensed and incubated for 2 h at room temperature. After that, 50 µL of biotinylated canine INSL3 (2 ng/mL in assay buffer) was added and incubated for a further 1 h. The biotinylated canine INSL3 was synthesized by the same procedure used for the biotinylated bovine INSL3 [14]. The wells were then washed three times with saline containing 0.05% Tween 20 and incubated for 30 min with horseradish peroxidase-labeled streptavidin (KPL, Gaithersburg, MD; 100 ng/mL in assay buffer). The wells were then again washed three times with saline containing 0.05% Tween 20 and incubated for another 30 min at room temperature with 100 µL substrate solution.
containing 3,3',5,5'-tetramethylbenzidine (TMB). The reaction was stopped by adding 50 μL of 2 M sulfuric acid, and the optical density was measured at 450 nm using an xMark microplate absorbance spectrophotometer (Bio-Rad Laboratories). The assay detection range was from 0.05 to 10 ng/mL. The intra- and inter-assay coefficients of variation were 14.7% and 16.2%, respectively. The hormonal specificity of the anti-bovine INSL3 antibody (2-8F) was validated previously [14]. The INSL3 peptide concentrations for each sample were normalized by protein amount in the homogenate. The protein amount was measured by BCA Protein Assay Reagent Kit (Thermo Scientific, Rockford, IL). Total amount of INSL3 peptide per testis was calculated from the data of INSL3 concentration and testicular weight.

2.6. Immunohistochemistry

Testicular tissues were examined by immunohistochemistry to check the expression of INSL3 peptide. Briefly, sections were cut from OCT-embedded tissue using a Cryostat (Leica CM1510S, Leica Microsystems, Wetzlar, Germany) at 7 μm and attached on glass slides (Platinum, Matsunami Glass, Osaka, Japan) treated with an anti-stripping reagent. The slide glasses were then immersed in a bottle containing PBS for washing, and the washing was repeated by transferring the slides into second and third
washing bottles.

ImmPRESS™ Reagent Kit Peroxidase Anti-Mouse Ig and the Peroxidase Substrate Kit DAB (Vector Laboratories, Burlingame, CA) were used for the immunohistochemistry. Each slide was then blocked with 250 μL of 2.5% normal horse serum and incubated for 20 min. After the blocking solution was discarded, the sections were incubated overnight with the primary antibody (Anti-bovine INSL3 antibody [2-8F]; 1: 1000 dilutions). After incubation with the primary antibody, the slides were incubated in 0.3% H₂O₂ for 30 min for quenching endogenous peroxidases. Thereafter, 350 μL secondary antibody (Anti-mouse Ig) was applied on slides and left to stand for 30 min. Finally, 380 μL of DAB solution was applied on the slides and the reaction was stopped after approx. 5–10 min. All incubations were carried out at room temperature in a humidified chamber except for those with primary antibody (4°C).

Following the incubation with primary or secondary antibody or 0.3% H₂O₂, the sections were washed (3 × 5 min) in 0.01 M PBS solution (pH 7.4). Staining with Hematoxylin was done for the same testes specimens used for immunohistochemistry in a different slide to check the cellular structures of the normal and retained testes including the presence/absence of sperm. The specificity of the staining with anti-bovine INSL3 antibody was confirmed in parallel sections by using assay buffer instead
of primary antibody, which was considered the negative control for the specificity of the
INSL3 immunostaining.

2.7. Data analyses

To evaluate the mRNA and peptide changes with the age, we categorized the
 normal dogs (6 mo–10 y; n=46) into pubertal (6 mo–1 y; n=19), post-pubertal (1–5 y;
n=17) and middle age (5–10 y; n=10). Samples obtained from pubertal and post-
pubertal ages (n=36) were used (samples from the middle age were excluded) for the
comparison among the normal, retained and cryptorchid scrotal testes groups, because
all of the cryptorchid dogs were within the age range from pubertal to post-pubertal age.

Immunohistochemistry was done in pubertal (n=3), post-pubertal (n=4) and middle-age
(normal dogs and cryptorchid dogs (n=5).

We also categorized the normal dogs’ breeds into four groups: (1) Toy Poodles
(n=12), (2) Miniature Dachshunds (n=10), (3) Chihuahuas (n=7), and (4) others (n=17),
to compare breed differences by using the total testicular weight per body weight as a
parameter. The normal dogs that were used to monitor age-related quantitative changes
of mRNAs and the INSL3 peptide were analyzed, not the dogs used for the
immunohistochemistry. We used the Chi-square test to identify any differences in the
breed distribution between the normal (n=56) and cryptorchid (n=22) dogs.

The evaluations of INSL3, RXFP2, LH receptor, 3β-HSD mRNAs and INSL3 peptide were performed by a two-way analysis of variance (ANOVA) using generalized linear (GENLIN) models of SPSS version 22 software (IBM, Somers, NY) to assess the effects of age and the testicular status of the animal (normal, retained testes or cryptorchid scrotal testes). Differences in mRNAs and peptides among the various age groups were compared using pairwise comparisons of the GENLIN procedure by the least significant difference (LSD) post hoc test. Data are expressed as mean ± SEM. Differences were considered significant at P<0.05.

3. Results

The mean testicular weight increased significantly (P<0.05) from the pubertal (2.40 ± 0.40 g) to the post-pubertal age (3.46 ± 0.29 g), and did not change from the post-pubertal to the middle-age (3.72 ± 0.47 g) dogs. The testicular weight of the retained testes of the cryptorchid dogs (0.56 ± 0.06 g) was much lower (P<0.01) than the scrotal testes (2.02 ± 0.41 g) of the unilateral cryptorchid dogs and the normal testes (2.90 ± 0.26 g) of the normal dogs. The testicular weight of the scrotal testes tended to be lower (P=0.07) than that of the normal testes. There was no significant difference in
the testicular weight per body weight values among the various breeds (Toy Poodles, 1.09 ± 0.11; Miniature Dachshunds 1.48 ± 0.19; Chihuahuas 1.06 ± 0.21; others 1.20 ± 0.08) of normal dogs used for the analyses of age-related changes of mRNAs and INSL3 peptide concentrations. The distributions of breeds were also the same between the normal and cryptorchid dogs (data not shown).

The INSL3 mRNA concentrations decreased (P<0.05) from pubertal to post-pubertal and from post-pubertal to middle age (Fig 1A), whereas the INSL3 peptide concentrations did not differ significantly among the age groups (Fig. 1B). Total amount of INSL3 mRNA per testis tended to decrease (P=0.05) from pubertal to post-pubertal age and decreased (P<0.05) from pubertal to middle age (Fig. 1E). However, total amount of INSL3 peptide per testis did not differ significantly among the age groups (Fig. 1F). The RXFP2 mRNA concentrations increased significantly (P<0.01) from pubertal to post-pubertal age, but there was no difference between post-pubertal and middle age (Fig. 1C). Total amount of RXFP2 mRNA per testis did not change among the age groups (data not shown).

The LH receptor mRNA concentrations did not differ between pubertal and post-pubertal age in the normal dogs, but they decreased significantly (P<0.01) from pubertal to middle age (Fig. 1D). The concentrations of 3β-HSD mRNA did not differ
significantly among age groups in normal dogs (data not shown). Total amount of LH receptor and 3β-HSD mRNAs per testis did not change among the age groups (data not shown).

The INSL3 mRNA concentrations were significantly higher (P<0.01) in the retained testes of the cryptorchid dogs compared to the normal testes of the normal dogs and the scrotal testes of the unilateral cryptorchid dogs (Fig. 2A). A very similar INSL3 mRNA concentration was observed between the normal testes of normal dogs and the scrotal testes of unilateral cryptorchid dogs (Fig. 2A). The total amount of INSL3 mRNA per retained testis was significantly lower (P<0.01) than that per normal testis and did not differ significantly from that per cryptorchid scrotal testis (Fig. 2E). The INSL3 peptide concentrations in the retained testes were significantly higher (P<0.05) than those in the cryptorchid scrotal testes and tended to be higher (P=0.08) than those in the normal testes (Fig. 2B). The INSL3 peptide concentrations for the scrotal testes of the unilateral cryptorchid dogs did not differ from the normal testes of the normal dogs (Fig. 2B). The total amount of INSL3 peptide per retained testis was significantly lower (P<0.01) than that per normal testis, but did not differ significantly from that per cryptorchid scrotal testis (Fig. 2F). The total amount of INSL3 peptide per cryptorchid scrotal testis tended to be lower (P=0.06) than that per normal testis (Fig. 2F).
The RXFP2 mRNA concentrations were almost negligible in the retained testes, and were much lower (P<0.001) than those in the normal testes (Fig. 2C). A similar concentration of RXFP2 mRNA was observed between the normal testes of the normal dogs and the scrotal testes of the unilateral cryptorchid dogs (Fig. 2C). The total amount of RXFP2 mRNA per retained testis was almost nil and much lower than that per normal testis (P<0.001) and cryptorchid scrotal testis (P<0.05; Fig. 2G). The total amount of RXFP2 mRNA per cryptorchid scrotal testis tended to be lower (P=0.08) than that per normal testis (Fig. 2G).

The LH receptor mRNA concentrations did not differ between the normal and cryptorchid dogs (data not shown). Total amount of LH receptor mRNA per retained testis was lower (P<0.01) than that per normal testis, and tended to be lower (P=0.14) than that per cryptorchid scrotal testis (Fig. 2H). Significantly higher (P<0.01) concentrations of 3β-HSD mRNA were observed in the retained and scrotal testes of the cryptorchid dogs compared to the normal testes of the normal dogs (Fig. 2D). Total amount of 3β-HSD mRNA per retained testis did not differ significantly among normal, retained and cryptorchid scrotal testes (data not shown).

We performed immunohistochemistry to examine the specific cell type(s) that shows INSL3 peptide expression in various age groups of normal testes of normal dogs.
and retained testes of cryptorchid dogs. Only Leydig cells of both the normal (pubertal, post-pubertal and middle age) and retained testes were immune-reactive to INSL3 antibody (shown in supplemental Fig. 1). The size of the seminiferous tubules per testicular area seemed to increase from pubertal to post-pubertal and middle age. The intensity of staining for INSL3 was clearly stronger in the Leydig cells of the retained testes compared to those of the normal testes in all age categories. No other testicular cell showed any immune reaction for INSL3 antibody (shown in supplemental Fig. 1). When the primary antibody was omitted, no immunostaining was observed (shown in supplemental Fig. 1). The Hematoxylin staining revealed the presence of sperm inside seminiferous tubules in the testes of the normal dogs (pubertal, post-pubertal and middle age) but the absence of sperm in the retained testes of the cryptorchid dogs (data not shown).

4. Discussion

It was reported that in rodents, INSL3 has pivotal roles in testicular descent in the fetal period [5, 6]. A role of INSL3 in the reproductive organs of domestic animals after puberty has rarely been reported. The changes of testicular INSL3 and its receptor, RXFP2, in pubertal and post-pubertal normal male animals have not yet been
elucidated, and a quantitative comparison of the testicular INSL3-receptor system between cryptorchid and normal animals has not been reported. In this study, we examined the gene expressions of INSL3 and RXFP2 in testes during puberty, post-puberty and middle age in normal dogs to elucidate the changing pattern of these genes’ expression with age and sexual maturity. We also compared the INSL3 and RXFP2 gene expressions in retained and scrotal testes of cryptorchid dogs with those of normal testes of normal dogs. This is apparently the first study regarding quantitative changes of testicular INSL3 and RXFP2 gene expression with age and sexual maturity and the comparison of these expressions between normal and cryptorchid dogs.

The present results revealed that total amount of INSL3 mRNA per testis decreased by aging in normal dogs despite increase of testicular weight, although the amount of INSL3 peptide per testis did not change significantly during the same ages. These results may indicate that the transcriptional activity of the gene encoding INSL3 in canine testes is reduced by the aging, but such a change is not reflected in the peptide content. It was suggested that INSL3 concentrations in peripheral blood are higher in the pubertal age and decline in the post-pubertal age in male dogs [15]. The change of INSL3 mRNA amount per testis from puberty to middle age observed in the present study, but not of the peptide, is likely to correspond to those of INSL3 concentrations in
blood. The reasons for the inconsistency of changes between testicular INSL3 mRNA and the peptide amount around canine puberty observed in our study are unknown. In male rats, the INSL3 concentrations in plasma transiently increased during puberty and decreased after puberty [13]. Testicular INSL3 mRNA concentrations in rats of advanced age (22–24 mo) were reduced compared to post-pubertal age (3 mo) [38]. In the present study, we did not examine histological changes of testicular cellular components including Leydig cells and various stages of germ cells during aging in the same samples which were measured for mRNAs and INSL3 peptide. Clearly, further studies are required to elucidate changes of INSL3 expression level per Leydig cell basis during puberty and aging in dogs.

We found higher mRNA and slightly increased peptide concentrations of INSL3 in the retained testes of the cryptorchid dogs compared to the normal testes of the normal dogs in the present study. Our immunohistochemistry data also showed that the areas occupied by INSL3-producing Leydig cells per a certain area of testicular tissue seem larger in the retained testes than in the normal testes, but we did not perform quantitative analyses of INSL3-producing Leydig cells in normal and retained testes in this study. However, total amounts of INSL3 mRNA and peptide per testis were reduced in the retained testes relative to normal testes due to much smaller size of the former,
suggesting that the canine retained testis may produce lower INSL3 as a whole testis. It has been suggested that INSL3 secretion in bilateral cryptorchid dogs is reduced compared to the normal and unilateral cryptorchid dogs [15]. The current study also shows that concentrations and total amount of INSL3 mRNA and peptide are similar between the scrotal testes of unilateral cryptorchid dogs and the normal testes of normal dogs. These results may be accorded with the previous findings that plasma INSL3 concentrations are similar between normal and unilateral cryptorchid dogs [15].

The present study provides findings that the gene expression of RXFP2 is almost disappeared in canine retained testes at both of per unit-weight basis and per whole-testis basis, in marked contrast to the higher expression of the receptor in normal testes. A previous histological examination also showed a lack of RXFP2 immunoreactivity in the genital tracts of cryptorchid canine testes [21]. Thus, it is likely in cryptorchid dogs that the substantial amount of INSL3 secreted in a retained testis cannot transduce its signal to cells within the testis, although we did not measure protein levels of RXFP2 receptor.

We speculate that the drastic reduction of RXFP2 mRNA in the retained testes may be caused mainly by the absence of advanced stages of germ cells that express RXFP2, due to impaired spermatogenesis [7, 19, 20]. It is also plausible that the down-
regulation of RXFP2 in Leydig cells by an autocrine mechanism [39, 40] with high or substantial concentrations of INSL3 could partly contribute to the loss of RXFP2 gene expression since relatively plenty of Leydig cells exist in the retained testes. It remains to be determined in future studies whether the down-regulation of RXFP2 occurs in Leydig cells in canine retained testes. We observed that the scrotal testes of unilateral cryptorchid dogs exhibit RXFP2 expression similar to that of the normal testes of normal dogs, implying that the lack of the receptor gene expression in the retained testes probably occurs as a consequence of — not as a cause of — the retention of the testes.

In addition to INSL3 and RXFP2, we analyzed the gene expression of LH receptor and 3β-HSD, which are also known as markers of Leydig cells [3, 27–29], during the course of puberty in the testes of the normal and cryptorchid dogs. Our findings revealed that mRNAs for both LH receptor and 3β-HSD showed differential dynamics compared with INSL3 mRNA during puberty and in the testes of the cryptorchid dogs. We speculate that the regulatory mechanisms for the gene expressions of these three markers for Leydig cells differ. It is not clear why the concentrations of 3β-HSD mRNA were increased not only in the retained but also in the scrotal testes of unilateral cryptorchid dogs in the present study. There could be a mechanism in unilateral cryptorchid dogs in which a retained testis may affect the function of the other
scrotal testis through substances secreted from the retained testis [34]. The region of

canine LH receptor mRNA selected for the real-time PCR in this study is known to
encode the receptor protein, but the transcript may slightly include splicing variants
which encode non-functional LH receptors. Thus it should be noted that not all of the
mRNA would be expressed as the functional LH receptor.

In conclusion, higher INSL3 mRNA per unit-weight basis and clear staining of
Leydig cells for INSL3 peptide in the retained testes of cryptorchid dogs indicate the
substantial expression of INSL3 in Leydig cells of the retained testes. However, smaller
amount of INSL3 is likely to be produced per a whole retained testis due to its
diminutive size. Also the present study reveals that RXFP2 gene expression is lost in the
retained testes, but occurs normally in cryptorchid scrotal testes.

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References


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Table 1. Oligonucleotide sequences of the primers used for real-time PCR, their location and the product sizes expected in canines

<table>
<thead>
<tr>
<th>Primer</th>
<th>Primer sequence (5’–3’)</th>
<th>Location</th>
<th>Product length (bp)</th>
</tr>
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<tbody>
<tr>
<td>INSL3</td>
<td>F: GGGGGCCCGCGCTGGTCCTC</td>
<td>145–164</td>
<td>181</td>
</tr>
<tr>
<td></td>
<td>R: CAGCTGCTCGCCGGTGTTGTTGATG</td>
<td>325–301</td>
<td></td>
</tr>
<tr>
<td>RXFP2</td>
<td>F: CAACTCACGCTACATCCATCAAAT</td>
<td>1292–1316</td>
<td>190</td>
</tr>
<tr>
<td></td>
<td>R: AGGACGGACACTTCAGTAGACAGC</td>
<td>1481–1458</td>
<td></td>
</tr>
<tr>
<td>LH receptor</td>
<td>F: TGTGTTGCCCTTCATCATCATTGT</td>
<td>1632–1655</td>
<td>346</td>
</tr>
<tr>
<td></td>
<td>R: AAGTTCAAGCCCGACGTTTACAGC</td>
<td>1977–1955</td>
<td></td>
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<tr>
<td>3β-HSD</td>
<td>F: CAGAATGCCACGAAGAAGAG</td>
<td>541–561</td>
<td>259</td>
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<td></td>
<td>R: AGACGGGGTTGACTATGGAGAA</td>
<td>799–778</td>
<td></td>
</tr>
<tr>
<td>18S rRNA</td>
<td>F: TGGTTGATCCTGCCGCTAGCA</td>
<td>5–25</td>
<td>96</td>
</tr>
<tr>
<td></td>
<td>R: ATGAGCCATTCGCAGTTTAC</td>
<td>100–79</td>
<td></td>
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Figure legends

**Fig. 1.** Changes in testicular concentrations of INSL3 mRNA (A), INSL3 peptide (B), RXFP2 mRNA (C), LH receptor mRNA (D), and total amount per testis of INSL3 mRNA (E) and INSL3 peptide (F) in various age groups of normal male dogs. Results are shown for pubertal (6–12 mo, n=19), post-pubertal (1–5 y, n=17) and middle age (5–10 y, n=10). Data are mean ± SEM. Values without a common superscript differed significantly for A and E (P<0.05), and for C and D (P<0.01).

**Fig. 2.** Testicular concentrations of INSL3 mRNA (A), INSL3 peptide (B), RXFP2 mRNA (C), 3β-HSD mRNA (D), and total amount per testis of INSL3 mRNA (E), INSL3 peptide (F), Rxfp2 mRNA (G) and LH receptor mRNA (H) in normal (n=36), retained (n=19) and scrotal testes (n=11). Data are mean ± SEM. Values without a common superscript differed significantly for B (P<0.05), for A, D, E, F and H (P<0.01) and for C and G (P<0.001).
Fig. 1
Fig. 2
Supplemental data Fig. 1
Supplemental data

Fig. 1. Representative photomicrographs of the immunohistochemical staining of INSL3 peptide (brown staining) in canine normal (A, pubertal; C, post-pubertal; E, middle age) and retained (G) testes. In both the normal and retained testes, only testicular Leydig cells (black arrows) showed INSL3 immunolabeling. The staining intensity was stronger in the retained testes compared to the normal testes in all age groups. When the primary antibody was omitted, immunolabeling was not observed in the normal (B, pubertal; D, post-pubertal; F, middle age) or retained (H) testes.